Master of Science in Molecular Medicine

Programme Code: MSMOLMED
Web page: www.ntnu.edu/studies/msmolmed

This programme description is valid for students admitted in the academic year 2016/2017.

Introduction
The field of Molecular Medicine is often referred to as “tomorrow’s medicine”. It aims to provide a molecular understanding of how normal cellular processes change, fail or are destroyed by disease.

The mapping of the human genome in 2003 was a turning point, and our knowledge and understanding of molecules in living organisms are advancing at a fast rate. Modern technologies such as high-throughput analyses (sequencing, microarray and proteomics) enable us to study thousands of genes and proteins simultaneously. This provides the foundation for a totally new understanding of biological systems and generates fresh hypotheses about the importance of genes and proteins for different diseases.

The MSc in Molecular Medicine is administered by the Department of Laboratory Medicine, Children’s and Women’s Health at the Faculty of Medicine.

Learning Objectives
The graduated candidate should be able to:

- demonstrate a strong background in molecular medicine (i.e. molecular/cell biology relevant to medical applications) and have practical skills relevant for the field;
- describe the organization of the human genome and its functional regulation (i.e. replication, gene expression, genome maintenance, and signal transduction principles);
- describe the impact of genes, inheritance and environment on disease, and understand how normal cellular processes change, fail or are destroyed by disease development, in particular for common diseases such as cancer, diabetes, and heart disease;
- explain principles of molecular diagnostics and advantages/limitations of its applications;
- recognize and explain current strategies and state-of-the-art approaches within functional genomics;
- collect relevant background information about topics within molecular medicine;
- present, evaluate and discuss scientific results in English (orally and in writing);
- reflect upon the existence of ethical aspects, sound experimental approaches and scientific thinking.
- discuss and solve relevant cases or problems in international teams/groups

Target Groups and Admission Requirements
Admission requirements to the MSc in Molecular Medicine is a bachelor’s degree (or an equivalent 3-year education) in biology, biomedical science, biotechnology, chemistry or similar, with an average grade of C or higher. A solid background in cell- and molecular biology is highly recommended within the bachelor’s degree.

International applicants need to submit proof of English proficiency (TOEFL, IELTS, APIEL or University of Cambridge test). More details about the language requirements are available at www.ntnu.edu/studies/lancourses/languagerequirements. Applicants who are not citizens of the European Union (EU) or the European Economic Area (EEA) need to provide a financial guarantee to get a residence permit in Norway.
Teaching Methods, Learning Activities and Student Social Activities

In 2005 the Laboratory Centre opened at Øya campus in Trondheim. In this building students get to work in high-tech laboratory environments side by side with researchers both from NTNU and St. Olav’s Hospital.

The teaching methods and learning activities include lectures, colloquiums, problem-based learning (PBL), team-based learning (TBL), seminars, demonstrations, excursions, practical training, self-tuition, and independent work. During the work with the master’s thesis the student will do research in our well-equipped laboratories.

SOMA is the master’s students’ own social student organization. SOMA has various activities during the semesters, including welcome parties and other activities for new students, excursions, courses and much more. For more information, visit SOMA’s blog: http://somantnu.blogspot.com

Compulsory HSE Training

All master’s students must participate in compulsory Health, Safety and Environment (HSE) training. This includes a HSE lecture and a fire protection course, both held in the first two weeks of the semester. When these activities have been completed, the student must pass an electronic test. This is to be done by 1 September 2016. If the student fails to do so, the access card to the campus/hospital buildings will be withdrawn.

Programme Structure

The MSc is a two-year, full-time programme of study starting in the autumn semester. There are two main components:

- Master's thesis (60 credits)
- Theoretical and methodological courses (totalling 60 credits). Two courses, making up 22.5 credits, are compulsory. The remaining courses, adding up to 37.5 credits, are selected from lists of electives. Ideally, electives should be linked to the topic of the master's thesis.

There are two lists of elective courses (see below). Two courses must be selected from ‘Electives 1’. The remaining elective courses can be chosen from both ‘Electives 1’ and ‘Electives 2’. Additional relevant courses may be taken at NTNU or other educational institutions subject to the approval of the Faculty of Medicine.

A master’s thesis agreement, including a project description, must be submitted by 15 March in the second semester. Potential projects will be presented in advance.

Master’s Thesis

<table>
<thead>
<tr>
<th>Course Code</th>
<th>Course Name</th>
<th>Credits</th>
</tr>
</thead>
<tbody>
<tr>
<td>MOL3901</td>
<td>Thesis in Molecular Medicine</td>
<td>60</td>
</tr>
</tbody>
</table>

Compulsory Courses

<table>
<thead>
<tr>
<th>Course Code</th>
<th>Course Name</th>
<th>Credits</th>
<th>Semester</th>
</tr>
</thead>
<tbody>
<tr>
<td>MOL3100</td>
<td>Introduction to Molecular Medicine with Project</td>
<td>15</td>
<td>Autumn</td>
</tr>
<tr>
<td>EiT</td>
<td>Experts in Teamwork – Interdisciplinary Project</td>
<td>7.5</td>
<td>Spring</td>
</tr>
</tbody>
</table>
### Electives 1

<table>
<thead>
<tr>
<th>Course Code</th>
<th>Course Title</th>
<th>Credits</th>
<th>Semester</th>
</tr>
</thead>
<tbody>
<tr>
<td>BI3016</td>
<td>Molecular Cell Biology</td>
<td>7.5</td>
<td>Autumn</td>
</tr>
<tr>
<td>MOL3001</td>
<td>Medical Genetics</td>
<td>7.5</td>
<td>Spring</td>
</tr>
<tr>
<td>MOL3005</td>
<td>Immunology</td>
<td>7.5</td>
<td>Autumn</td>
</tr>
<tr>
<td>MOL3007</td>
<td>Functional Genomics</td>
<td>7.5</td>
<td>Spring</td>
</tr>
</tbody>
</table>

### Electives 2

<table>
<thead>
<tr>
<th>Course Code</th>
<th>Course Title</th>
<th>Credits</th>
<th>Semester</th>
</tr>
</thead>
<tbody>
<tr>
<td>BI3013</td>
<td>Experimental Cell and Molecular Biology</td>
<td>7.5</td>
<td>Autumn</td>
</tr>
<tr>
<td>BI3018</td>
<td>Patenting and Commercialization of Biotech and Medtech Inventions</td>
<td>7.5</td>
<td>Autumn</td>
</tr>
<tr>
<td>KLH3100</td>
<td>Introduction to Medical Statistics</td>
<td>7.5</td>
<td>Spring</td>
</tr>
<tr>
<td>MOL3003</td>
<td>Molecular Medical Microbiology with Laboratory Work</td>
<td>7.5</td>
<td>Autumn</td>
</tr>
<tr>
<td>MOL3009</td>
<td>Biobanking</td>
<td>7.5</td>
<td>Autumn</td>
</tr>
<tr>
<td>MOL3010</td>
<td>Animal Cell Culture</td>
<td>7.5</td>
<td>Autumn</td>
</tr>
<tr>
<td>MOL3014</td>
<td>Nanomedicine I – Bioanalysis</td>
<td>7.5</td>
<td>Autumn</td>
</tr>
<tr>
<td>MOL3015</td>
<td>Nanomedicine II – Therapy</td>
<td>7.5</td>
<td>Spring</td>
</tr>
<tr>
<td>MOL3018</td>
<td>Medical Toxicology</td>
<td>7.5</td>
<td>Spring</td>
</tr>
<tr>
<td>MOL3020</td>
<td>Virology</td>
<td>7.5</td>
<td>Spring</td>
</tr>
<tr>
<td>MOL3021</td>
<td>Bioinformatics – Applied Project</td>
<td>7.5</td>
<td>Spring</td>
</tr>
<tr>
<td>MOL3022</td>
<td>Bioinformatics – Method Oriented Project</td>
<td>7.5</td>
<td>Spring</td>
</tr>
<tr>
<td>MOL3023</td>
<td>Molecular Medical Microbiology with Essay</td>
<td>7.5</td>
<td>Autumn</td>
</tr>
<tr>
<td>NEVR8014</td>
<td>Laboratory Animal Science for Researchers</td>
<td>7.5</td>
<td>Autumn</td>
</tr>
</tbody>
</table>

Some of the elective 2 courses may be cancelled if few students register for the examination.

Experts in Teamwork (EiT) is taught intensively in January in the first academic year. Read more about EiT at www.ntnu.edu/eit

### Model of the MSc Programme (Example)

<table>
<thead>
<tr>
<th>Year 1</th>
<th>Year 2</th>
</tr>
</thead>
<tbody>
<tr>
<td>1st semester (autumn)</td>
<td>2nd semester (spring)</td>
</tr>
<tr>
<td>Introduction to Molecular Medicine with Project</td>
<td>Experts in Teamwork</td>
</tr>
<tr>
<td>Elective course</td>
<td>Elective course</td>
</tr>
<tr>
<td>Elective course</td>
<td>Elective course</td>
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</tbody>
</table>

Please note that this is only a suggestion. The student can choose to start with the thesis already in the first year and postpone one or more of the elective courses to the second year.

The student must have passed all examinations in compulsory and elective courses before he/she can submit the thesis.
Course Descriptions

<table>
<thead>
<tr>
<th>BI3013</th>
<th>Experimental Cell and Molecular Biology</th>
</tr>
</thead>
<tbody>
<tr>
<td>Credits:</td>
<td>7.5</td>
</tr>
<tr>
<td>Period:</td>
<td>Autumn</td>
</tr>
<tr>
<td>Teaching methods:</td>
<td>Laboratory course / demonstrations (40 hours, compulsory)</td>
</tr>
<tr>
<td></td>
<td>Lectures (20 hours, compulsory)</td>
</tr>
<tr>
<td>Compulsory activities:</td>
<td>Laboratory course / demonstrations</td>
</tr>
<tr>
<td></td>
<td>Approved report</td>
</tr>
<tr>
<td>Mode of assessment:</td>
<td>4-hour written examination</td>
</tr>
<tr>
<td></td>
<td>Letter grades (A-F)</td>
</tr>
<tr>
<td>Credit reductions due to overlapping courses:</td>
<td>MNKBI313: 7.5 credits</td>
</tr>
<tr>
<td>Host department:</td>
<td>Department of Biology</td>
</tr>
<tr>
<td>Course coordinator:</td>
<td>Associate Professor Per Winge</td>
</tr>
</tbody>
</table>

BI3013 has restricted admission. Please contact the Department of Biology if you are interested.

Learning outcome
The aim of the course is to introduce basic methods in cell- and molecular biology. The course includes practical exercises in modern experimental techniques and instruments, and also training in literature search and the use of Internet. Selected analytical methods will be presented and tested. The course also includes analyses of problems and artefacts that generally occur in biological samples examined using chemical and biological analyses.

Academic content
On completion of the course students should be familiar with basic methods in cell- and molecular biology. Students should also be able to demonstrate knowledge of how to use modern experimental techniques and instruments.

<table>
<thead>
<tr>
<th>BI3016</th>
<th>Molecular Cell Biology</th>
</tr>
</thead>
<tbody>
<tr>
<td>Credits:</td>
<td>7.5</td>
</tr>
<tr>
<td>Period:</td>
<td>Autumn</td>
</tr>
<tr>
<td>Teaching methods:</td>
<td>Lectures (26 hours) and seminars (24 hours, mandatory)</td>
</tr>
<tr>
<td>Mode of assessment:</td>
<td>4-hour written examination</td>
</tr>
<tr>
<td>Credit reductions due to overlapping courses:</td>
<td>MNKBI316 7.5 credits</td>
</tr>
<tr>
<td>Host department:</td>
<td>Department of Biology</td>
</tr>
<tr>
<td>Course coordinator:</td>
<td>Associate Professor Per Winge</td>
</tr>
</tbody>
</table>

Learning outcome
On completion of the course students should have an understanding of cell biology mechanisms on a molecular level, and of the regulation of such mechanisms.

Academic content
Subjects covered include: Apoptose/necrose mechanisms; Kinases/phosphatases classification and regulation; Transcription factors, classification and regulation; Lipid mediators, regulation and function mechanisms; DNA repair mechanisms. Syllabus will mainly be based on research- and review articles.
BI3018 | Patenting and Commercialization of Biotech and Medtech Inventions

Credits: 7.5
Period: Spring

Teaching methods: The course is held intensively during one week during the months March / April. Lectures and case-based work in groups are repeated for every theme in the course. Oral presentation of work in groups by students. Written assignments are to be submitted two/three weeks after completion of the intensive part of the course. These are performed in groups. Submission written project assignment.

Recommended previous knowledge: Target group: Master’s and PhD students, Tech Trans personnel, Biotech/Medtech staff
Required previous knowledge: Bachelor’s degree or equivalent.

Mode of assessment: Report
Letter grades (A-F)
Host department: Department of Biology
Course coordinator: Professor Berit Johansen

Learning outcome

Knowledge: The candidate shall have knowledge about:
- aspects involved in transforming a research project to commercial product
- IP management;
- patenting; basics, process, national/international law, regulations, practising, similarities/differences;
- scientific versus commercial aspects on patenting strategy/IP evaluations;
- processes involved in transforming a research product to a clinical product;
- models for sale of IP, licensing versus sale;
- business development: IP, business plan, coworkers, financing.

Skills: The candidate can:
- identify and describe the different processes important for conservation of intellectual property of an invention and how to commercialize;
- identify and describe criteria and processes for sale of IP, including business development.

General competence: The candidate can:
- identify and explain principles in processes regulating protection and sale of IP.

Academic content

Topics that will be covered in the course include:
- Patenting: Principles, process, national/international laws, regulations and practice, similarities/differences between European and US patenting laws and practise.
- IPR strategies: Scientific/commercial aspects, how to develop an IP strategy to accelerate the innovation process and to safeguard IP investments, mastering freedom to operate in the Biotech/MedTech industry, Patent litigations, infringements and enforcements.
- Licensing: Models and negotiation strategies.
- Clinical testing: Design, implementation, analysis and presentation of clinical trials, adaptive clinical trial designs.
- Bio-tech/Med-tech business development: Strategy and organization when transferring a scientific idea into a commercial product/business, business plan development, product pipeline analysis, market analysis, market potential prediction, alliance structures and negotiation conditions, capital capture (pre-seed, seed, VC).
Learning outcome

After completing the course KLH3100, the student should be able to

- select appropriate summary statistics and graphical displays for describing data for continuous and categorical variables in an empirical data set;
- describe and apply statistical methods for comparing a mean value or a proportion in one sample to a reference value, and for comparing mean values or proportions in two independent or paired samples;
- describe and apply the methods of correlation and simple linear regression for identifying associations or relationships between two continuous variables, and methods for evaluating agreement in repeated measures for continuous and categorical data;
- select the appropriate statistical method for analyzing a specific research question, study design and data set;
- perform statistical analyses to an empirical data set by means of a statistical software package (SPSS);
- interpret and present the results from statistical analyses, and critically evaluate the validity of the results in light of the assumptions for the chosen method.

Academic content

- Introduction to the statistical software package SPSS.
- Descriptive statistic for continuous and categorical variables (measures of location and spread, frequency tables, graphical display), probability and probability distributions, estimation, hypothesis testing, one- and two-sample tests on mean values (Student T-test), non-parametric tests (Wilcoxon signed-rank test and Mann-Whitney U test), tests on differences in proportions (cross-table analysis; chi-square test and McNemar’s test, Fisher’s exact test), correlation and simple linear regression, methods for assessing agreement (Kappa coefficient, Bland-Altman plot).
### MOL3001 Medical Genetics

<table>
<thead>
<tr>
<th><strong>Credits:</strong></th>
<th>7.5</th>
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</thead>
<tbody>
<tr>
<td><strong>Period:</strong></td>
<td>Spring</td>
</tr>
<tr>
<td><strong>Teaching methods:</strong></td>
<td>Lectures, student presentations, laboratory course and alternative methods of teaching. The lectures and the exam will be in English. If few candidates, alternative exam arrangements may be used.</td>
</tr>
<tr>
<td><strong>Recommended previous knowledge:</strong></td>
<td>Biochemistry and basic genetics</td>
</tr>
<tr>
<td><strong>Compulsory activities:</strong></td>
<td>Laboratory course</td>
</tr>
<tr>
<td><strong>Mode of assessment:</strong></td>
<td>4-hour written examination</td>
</tr>
<tr>
<td><strong>Letter grades (A-F):</strong></td>
<td></td>
</tr>
<tr>
<td><strong>Host department:</strong></td>
<td>Department of Laboratory Medicine, Children’s and Women’s Health</td>
</tr>
<tr>
<td><strong>Course coordinator:</strong></td>
<td>Associate Professor Wenche Sjursen</td>
</tr>
</tbody>
</table>

### Learning outcome

After completing the course MOL3001 the student is able to:

- describe central examples of monogenic, polygenic and chromosomal disorders;
- recognise patterns of mendelian inheritance of monogenic diseases, and explain genetic and biochemical mechanisms of some central monogenic disorders;
- describe and understand mechanisms underlying numerical and structural chromosomal aberrations and principles mediating chromosomal disease;
- describe what genetic counselling and risk assessment are;
- discuss bioethical issues in medical genetics;
- describe and understand central principles and examples of both sporadic and hereditary cancers;
- describe and understand principles for methods of genetic diagnosis, i.e. gene tests and cytogenetic methods;
- describe and understand principles and methods for gene mapping;
- calculate frequencies of genetic variants at individual and population based level.

### Academic content

The course will give an overview of mechanisms for development of genetic diseases. Topics include different patterns of inheritance, like dominant, recessive, autosomal and sex linked inheritance. Genetic diseases will be classified in single-gene, chromosomal and multifactorial disorders. It will be discussed how identification of genes and variants in the genome, including gene mapping, make it possible to understand how variation can lead to disease.
<table>
<thead>
<tr>
<th>MOL3003</th>
<th>Molecular Medical Microbiology with Laboratory Work</th>
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</thead>
<tbody>
<tr>
<td>Credits:</td>
<td>7.5</td>
</tr>
<tr>
<td>Period:</td>
<td>Autumn</td>
</tr>
<tr>
<td>Teaching methods:</td>
<td>Lectures, demonstrations and laboratory exercises, and a compulsory written assignment which counts 25% of the final grade. The language of teaching and examination is English. The course has restricted admission. Two-thirds of the seats are reserved for first-year master's students in molecular medicine. One-third of the seats are reserved for second-year master's students in molecular medicine and other master's students at NTNU. Timetables for courses at the Faculty of Medicine are available at <a href="https://timeplan.medisin.ntnu.no/timetable_show.php">https://timeplan.medisin.ntnu.no/timetable_show.php</a></td>
</tr>
<tr>
<td>Recommended previous knowledge:</td>
<td>Knowledge in microbiology, cell biology, biochemistry, and/or molecular biology</td>
</tr>
<tr>
<td>Compulsory activities:</td>
<td>Guided self study, including laboratory work</td>
</tr>
<tr>
<td>Mode of assessment:</td>
<td>Oral examination – 75% of the final grade</td>
</tr>
<tr>
<td></td>
<td>Written assignment – 25% of the final grade</td>
</tr>
<tr>
<td></td>
<td>Letter grades (A-F)</td>
</tr>
<tr>
<td>Credit reduction due to overlapping course:</td>
<td>MOL3023: 7.5 credits</td>
</tr>
<tr>
<td></td>
<td>MOL8011: 7.5 credits</td>
</tr>
<tr>
<td>Host department:</td>
<td>Department of Laboratory Medicine, Children’s and Women’s Health</td>
</tr>
<tr>
<td>Course coordinator:</td>
<td>Professor Jan Egil Afset</td>
</tr>
</tbody>
</table>

The course has restricted admission. Two-thirds of the seats are reserved for first-year master's students in molecular medicine. One-third of the seats are reserved for second-year master's students in molecular medicine and other master's students at NTNU.

**Learning outcome**

After completing the course MOL3003 the student is able to:

- demonstrate a strong general knowledge in molecular medical microbiology;
- have good knowledge of the PCR method including design, optimization and validation for detection, identification and quantification of microorganisms;
- demonstrate basic skills in performing PCR in the laboratory;
- discuss interpretation of PCR results and limitations of the method;
- demonstrate knowledge of other molecular methods for identification and quantification of microorganisms;
- demonstrate good knowledge of molecular methods for genotyping of microbial agents, and discuss the use of molecular epidemiological methods in the investigation of infectious diseases;
- describe the use of bioinformatics tools in molecular medical microbiology, and have basic skills in the use of such methods;
- describe and discuss the use of relevant quality control measures in the molecular medical microbiology laboratory.

**Academic content**

Molecular methods are important for the diagnosis of infectious diseases and for characterization of the microorganisms causing such infections. In this course the student will be introduced to the use of molecular genetic methods in the diagnosis of human pathogenic microorganisms, including methods for extraction of nucleic acids, qualitative and quantitative PCR and DNA sequencing. Principles for primer and probe design will be explained, as well as use of bioinformatics in the design of PCR assays. Methods for genotyping of microorganisms are important for the study of transmission and dissemination of infectious diseases in hospitals and in the community. A range of genotyping methods developed for this purpose will be presented and discussed.
MOL3005 Immunology

Credits: 7.5
Period: Autumn
Teaching methods: Lectures and colloquiums (not compulsory). The language of teaching is English. Timetable: https://timeplan.medisin.ntnu.no/timetable_show.php. Information will be communicated on It's learning.

Recommended previous knowledge: Basic knowledge within cell biology and biochemistry/molecular biology.

Mode of assessment: 4-hour written examination
Letter grades (A-F)

Credit reductions due to overlapping courses: BI2013: 7.5 credits
MNKBI213: 7.5 credits

Host department: Department of Laboratory Medicine, Children’s and Women’s Health

Course coordinator: Researcher Ingvild Bjellmo Johnsen

Learning outcome

After completing the course MOL3005 the student is able to:

- demonstrate the basic knowledge of immunological processes at a cellular and molecular level;
- define central immunological principles and concepts;
- outline, compare and contrast the key mechanisms and cellular players of innate and adaptive immunity and how they relate;
- understand the principles of central (antibody-based) immunological methods to an extent that he/she can set up a theoretical experiment;
- elucidate the genetic basis for immunological diversity and the generation of adaptive immune responses;
- outline key events and cellular players in antigen presentation, and how the nature of the antigen will shape resulting effector responses;
- identify the main mechanisms of inflammation;
- understand the principles governing vaccination and the mechanisms of protection against disease;
- understand how immunodeficiencies related to disease;
- understand and explain the basis of allergy and allergic diseases.

Academic content

The immune system governs defense against pathogens and is of importance for development of autoimmune diseases, allergy and cancer. The course discusses basic immunology including cellular and molecular processes that represents the human immune system. Subjects to be presented include cells and organs of the immune system, antigen, immunoglobulins and antibody diversity, molecular mechanisms of innate and adaptive immunity, the complement system, antigen presentation, cell-mediated effector responses, immunological techniques and select lecture on the immune system in health and disease.
MOL3007     Functional Genomics
Credits:     7.5
Period:     Spring
Teaching methods: Lectures, laboratory course. The lectures are held in the spring semester, and starts in early February. The language of instruction and examination is English. Timetable: https://timeplan.medisin.ntnu.no/timetable_show.php
Recommended previous knowledge: Basic skills in molecular biology and physiology.
Compulsory activity: Laboratory course
Mode of assessment: 4-hour written examination
Letter grades (A-F)
Host department: Department of Cancer Research and Molecular Medicine
Course coordinator: Professor Astrid Lægreid

Learning outcome
After completing the course MOL3007 the student is able to:

- describe what is meant by functional genomics and how this area of research contributes both to new basic biomedical knowledge and to new developments in biomedicine and biotechnology, including improved diagnostics and treatment of disease;
- describe and discuss how functional genomics contributes to systems biology and systems medicine;
- understand and discuss the interdependence of biomedicine/biotechnology, bioinformatics and bioethics within functional genomics;
- reflect on ethical and societal aspects of functional genomics translated to health care;
- explain how genetically modified organisms can contribute to functional genomics research;
- explain main principles of high throughput transcriptomic and genomic analysis by state of the art sequencing and microarray technology;
- explain mechanisms of gene regulation;
- explain the concepts of structural motifs and domains in proteins and methods used to assess these structures;
- explain main principles of central methods used in protein separation;
- explain main methods for ionisation of peptides and how mass analysis of peptides and fragments thereof can be used to identify proteins and their post-translational modifications;
- explain main principles of heterologous expression and imaging and its contribution to functional genomics research - explain main principles of bioinformatic tools used for –omics data analysis, biological background knowledge management and modeling.

Academic content
- Fundamental principles within functional genomics, emphasizing the transcriptome and proteome.
- Contribution of functional genomics to systems biology and systems medicine.
- Socio-ethical aspects of functional genomics in biomedicine and biotechnology, including perspectives on genetic risk information.
- Hypothesis generation/experimental design.
- Experimental model systems.
- Next generation HTP sequencing technology.
- Microarray-technology.
- Protein separation (2D-PAGE, 2D-LC).
- Mass spectrometry (MALDI-TOF, ESI-MS).
- Protein structure analysis (X-ray crystallography, NMR).
- Heterologous expression. Imaging.
### MOL3009 Biobanking

**Credits:** 7.5  
**Period:** Autumn

**Teaching methods:** Lectures, seminar, group exercises, laboratory exercise, excursion, semester project. The language of teaching and examination is English. If few students sign up, the course may be cancelled. If the number of students exceed a given number, alternative examination might be considered. Timetables for courses at the Faculty of Medicine are available at https://timeplan.medisin.ntnu.no/timetable_show.php

**Recommended previous knowledge:** Basic knowledge in medical genetics and cell biology

**Compulsory activities:** Excursion, laboratory exercise and project work

**Mode of assessment:** Oral examination  
Letter grades (A-F)

**Host department:** Department of Laboratory Medicine, Children’s and Women’s Health

**Course coordinators:** Professor Jostein Halgunset

### Learning outcome

After completing the course MOL3009 the student is able to:

- describe and explain universal and special features of the different types of biobanks;
- explain the different conditions and requirements that must be fulfilled with regard to operation and use of research biobanks;
- describe quality management, quality assurance, logistics and data management related to biobanks;
- discuss ethical issues and policy guidance regarding research involving human biological materials;
- outline the main features of laws and other directives which are relevant for biobanking;
- discuss the use and the potential utility values of biobanks;
- outline how to establish and operate a biobank;
- describe in detail the application process for the establishment of a research biobank - sketch the plan for a research project using human biological material.

### Academic content

This course will focus on the following topics:

- Biobanks: classifications, common and distinctive features of the different types of biobanks.
- National regulations and international conventions concerning biobanks, use of human biological materials and personal data.
- Research biobanks: formal and practical aspects of the establishment, operation and use
- Ethical aspects of utilization of human biological materials; The role of Research Ethics Committees
- Different types of consent as basis for biobanking; alternatives to consent
- Logistics and quality management; quality assurance and quality control of collection, storage, retrieval and use of samples
- Methods for analysis of human biological materials; assessment and interpretation of data
- Extraction and quality assessment of DNA and/or RNA from various sources
- Collection and storage of data; databases and data security
- Statistical and epidemiological methods in biobank related research
- The role of biobanks in health surveys like HUNT, Mother-Child etc.
- Research for the future: National and transnational genetic and epidemiological research collaboration
# MOL3010 Animal Cell Culture

<table>
<thead>
<tr>
<th>Credits:</th>
<th>7.5</th>
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</thead>
<tbody>
<tr>
<td>Period:</td>
<td>Autumn</td>
</tr>
<tr>
<td>Teaching methods:</td>
<td>Self-tuition. The language of the examination is English.</td>
</tr>
<tr>
<td>Recommended previous knowledge:</td>
<td>Basic knowledge in cell biology and biochemistry.</td>
</tr>
<tr>
<td>Mode of assessment:</td>
<td>Oral examination</td>
</tr>
<tr>
<td>Letter grades (A-F)</td>
<td></td>
</tr>
<tr>
<td>Host department:</td>
<td>Department of Laboratory Medicine, Children’s and Women’s Health</td>
</tr>
<tr>
<td>Course coordinator:</td>
<td>Post Doctor Caroline Hild Hakvåg Pettersen</td>
</tr>
</tbody>
</table>

*Please note that this course is based on self-tuition. It will not be given any lectures.*

## Learning outcome

After completing the course MOL3010 the student is able to:

- demonstrate knowledge of basic cell culture techniques;
- demonstrate knowledge of establishment of cell inlines and their maintenance;
- demonstrate knowledge on design and use the cell culture facilities;
- critically evaluate cell cultures constraints and possibilities as an in vitro model;
- discuss the advantages and limitations of primary cell culture compared to immortalized or transformed cell lines.

## Academic content

The course will focus on practical aspects of cell culture, like design and layout of the laboratory, aseptic technique, cloning and selection of specific cell types, contamination, methods for measuring viability and cytotoxicity, cell culture environment (substrate, gas phase, medium) and the culturing of specific cell types.

# MOL3014 Nanomedicine I - Bioanalysis

<table>
<thead>
<tr>
<th>Credits:</th>
<th>7.5</th>
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<tbody>
<tr>
<td>Period:</td>
<td>Autumn</td>
</tr>
<tr>
<td>Teaching methods:</td>
<td>The syllabus of the course is defined by the learning objectives. The course is based on lectures given by experienced researchers within each theme. The course includes a compulsory project providing an in-depth review of the primary litterature, which will account for 25 % of the final grade. There might be simple lab exercises dependent on number of students enrolled. The language of instruction is English. Timetable: <a href="https://timeplan.medisin.ntnu.no/timetable_show.php">https://timeplan.medisin.ntnu.no/timetable_show.php</a></td>
</tr>
<tr>
<td>Recommended previous knowledge:</td>
<td>Basic skills in molecular biology, cell biology, chemistry, physics. Most suited for students who have completed courses in basic molecular and cell biology.</td>
</tr>
<tr>
<td>Mode of assessment:</td>
<td>4-hour written examination – 75 % of the final grade</td>
</tr>
<tr>
<td></td>
<td>Exercise / Project – 25 % of the final grade</td>
</tr>
<tr>
<td></td>
<td>Letter grades (A-F)</td>
</tr>
<tr>
<td>Host department:</td>
<td>Department of Cancer Research and Molecular Medicine</td>
</tr>
<tr>
<td>Course coordinator:</td>
<td>Professor Øyvind Halaas</td>
</tr>
</tbody>
</table>

## Learning outcome

After completing the course MOL3014 the student is able to:

- understand how nanotechnology can be tailored and used for biomedical purposes;
- understand the problems and possibilities for analysis of proteins, nucleic acids and cells by micro fabricated devices and nanotechnological solutions;
- outline fabrication procedures and general considerations for microfluidics;
• understand how nano-relevant instruments such as focused ion beam scanning electron microscopes, atomic force microscopes and optical microscopes can be used in biomedicine;
• perform simple micro fabrication procedure;
• find, refer and consider relevant information.

Academic content
This course will cover fundamentals of bioanalysis and module integration for applications. In detail the course will contain:
• Advanced protein and DNA chemistry.
• Methods for quantification and identification of DNA/RNA and protein with focus on technical principles and emerging nanotechnologies.
• Use of imaging in nanoscale for biomedical research.
• Microfluidics.
• Principles for and construction of lab-on-a-chip and biosensors.
• Nanoneuroscience.

This course is focused on technology rather than biology.

<table>
<thead>
<tr>
<th>MOL3015</th>
<th>Nanomedicine II - Therapy</th>
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</thead>
<tbody>
<tr>
<td>Credits:</td>
<td>7,5</td>
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<tr>
<td>Period:</td>
<td>Spring</td>
</tr>
<tr>
<td>Teaching methods:</td>
<td>The syllabus of the course is defined by the learning objectives. The course is based on lectures given by experienced researchers within each theme. The course includes a compulsory project providing an in-depth review of the primary literature, which will account for 25 % of the final grade. The language of instruction is English. The lectures are held in the spring semester and start in early February. Timetable: <a href="https://timeplan.medisin.ntnu.no/timetable_show.php">https://timeplan.medisin.ntnu.no/timetable_show.php</a></td>
</tr>
<tr>
<td>Recommended previous knowledge:</td>
<td>Basic skills in molecular biology.</td>
</tr>
<tr>
<td>Mode of assessment:</td>
<td>4-hour written examination – 75 % of the final grade</td>
</tr>
<tr>
<td></td>
<td>Exercise / Project – 25 % of the final grade</td>
</tr>
<tr>
<td></td>
<td>Letter grades (A-F)</td>
</tr>
<tr>
<td>Host department:</td>
<td>Department of Cancer Research and Molecular Medicine</td>
</tr>
<tr>
<td>Course coordinator:</td>
<td>Professor Øyvind Halaas</td>
</tr>
</tbody>
</table>

Learning outcome
After completing the course MOL3015 the student is able to:
• understand how nanotechnological approaches can be used in biomedical therapies;
• understand biomaterials and interaction of biomaterials with cells, body fluids and tissues;
• understand basic stem cell biology and corresponding requirement for tissue engineering;
• understand the need, obstacles and solutions for polymeric, lipidous and solid nanosized drug delivery systems;
• understand the toxicological aspects of nanosized surfaces and particles;
• find, refer and evaluate available information.

Academic content
The course will introduce use of nanotechnology in therapy. In detail, the course will cover
• Clinical biomaterials, tissue regeneration, including stem cell technology, immunological limitations and encapsulation strategies.
• Methods and possibilities for drug discovery.
• Use and design of nanoparticles for gene therapy, drug delivery and drug targeting.
• Physiological, cellular and toxicological limitations for medical use of nanoparticles.
• Theranostics, the combined use of in vivo imaging/diagnostics and therapy.
• Ethical, legal and social aspects (ELSA) related to use of medical nanotechnology will be discussed.

A written report is included, where the student will choose a theme from the lectures, review the literature, describe current methods, consider and recommend use of emerging nanotechnologies in a therapeutic setting.

<table>
<thead>
<tr>
<th>MOL3018</th>
<th>Medical Toxicology</th>
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<tbody>
<tr>
<td>Credits:</td>
<td>7.5</td>
</tr>
<tr>
<td>Period:</td>
<td>Spring</td>
</tr>
<tr>
<td>Teaching methods:</td>
<td>Lectures. The language of instruction and examination is English. The course is taught in the spring semester, and starts in late January or early February. Timetable: <a href="https://timeplan.medisin.ntnu.no/timetable_show.php">https://timeplan.medisin.ntnu.no/timetable_show.php</a></td>
</tr>
<tr>
<td>Recommended previous knowledge:</td>
<td>Passed examinations in BI1001 and BI1004, or TBT4100 and TBT4105 (or similar courses).</td>
</tr>
<tr>
<td>Required previous knowledge:</td>
<td>Basic knowledge of physiology, chemistry, biochemistry and mathematics.</td>
</tr>
<tr>
<td>Mode of assessment:</td>
<td>4-hour written examination</td>
</tr>
<tr>
<td></td>
<td>Letter grades (A-F)</td>
</tr>
<tr>
<td>Credit reductions due to overlapping courses:</td>
<td>TOKS1010: 7.5 credits</td>
</tr>
<tr>
<td></td>
<td>TOKS3010: 7.5 credits</td>
</tr>
<tr>
<td>Host department:</td>
<td>Department of Cancer Research and Molecular Medicine</td>
</tr>
<tr>
<td>Course coordinator:</td>
<td>Associate Professor Bent Håvard Hellum</td>
</tr>
</tbody>
</table>

**Learning outcome**

After completing the course MOL3018, the student is able to:

• describe and explain toxicological mechanisms;
• perform simple analysis of how some chemicals might be a possible health hazard upon exposure;
• explain how certain xenobiotics in the environment and work can have toxic effects on central organs and organ systems in humans;
• collect relevant background data regarding toxicological problems.

**Academic content**

The course gives an introduction to general pharmacokinetic models. Liver, kidney, lung, the immunonervous system will be discussed as target organs for chemical toxicity. Groups of toxic agents and substances of abuse will also be included. Major weight will be put on available methods for risk assessment of human exposure to cancer and non-cancer agents.
**Learning outcome**

After completing the course MOL3020, the student is able to:

- demonstrate a general knowledge in molecular virology;
- describe elements of the viral life cycle;
- explain viral replication strategies;
- describe viral recognition mechanisms and immunological defense responses;
- discuss principles of virus pathogenesis;
- outline viral molecular epidemiology;
- explain vaccine strategies and mechanisms of antiviral drugs.

**Academic content**

The course will give an overview of medically important virus families, their replication strategies and mechanisms for development of viral infectious diseases.

Topics will include taxonomy, replication strategies, pathogenicity and transmission of viruses and, additionally, diagnosis, prevention and treatment of viral diseases. Antiviral immunity and viral immune-evasion will also be covered. Common human viral infections will be the main focus of the course, and emphasis will be put on virus-host interactions as a key to understanding the diversity of viruses and viral diseases.
**Learning outcome**

After completing the course MOL3021 the student is able to:

- explain function and usage of important bioinformatics tools, in particular for analyzes at the sequence level (genome, gene, RNA and protein);
- describe important formats for storage and exchange of biological information;
- describe content and use of important bioinformatics databases and web portals;
- use bioinformatics tools and databases to analyze relevant data from molecular biology;
- use relevant bioinformatics tools and databases in own projects.

**Academic content**

The course gives a practical introduction to important methods in bioinformatics, including sequence library searches, pairwise and multiple alignment, phylogenetic analysis, gene prediction, ontology-based analyses, motif discovery and structure prediction. The students will be able to test the methods on realistic problems through computer-based exercises. The students will also do a project where different methods are combined. An individual report from the project will be the basis for grading. The course is in particular targeted towards students with a background from molecular biology who wants to use bioinformatics in an applied context, for example for analysing own data.

<table>
<thead>
<tr>
<th>MOL3022</th>
<th>Bioinformatics – Method Oriented Project</th>
</tr>
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<tbody>
<tr>
<td>Credits:</td>
<td>7.5</td>
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<tr>
<td>Period:</td>
<td>Spring</td>
</tr>
<tr>
<td>Teaching methods:</td>
<td>Lectures and exercises (computer lab). The lectures are given intensively over two weeks in the spring semester, including work with the project assignment. The language of instruction is English. The timetable will be available at <a href="https://timeplan.medisin.ntnu.no/timetable_show.php">https://timeplan.medisin.ntnu.no/timetable_show.php</a></td>
</tr>
<tr>
<td>Recommended previous knowledge:</td>
<td>Basic knowledge in molecular biology, statistics and informatics.</td>
</tr>
<tr>
<td>Compulsory activity:</td>
<td>Exercises</td>
</tr>
<tr>
<td>Mode of assessment:</td>
<td>Assignment Letter grades (A-F)</td>
</tr>
<tr>
<td>Credit reduction due to overlapping course:</td>
<td>MOL3019: 7.5 credits MOL3021: 7.5 credits MTEK3001: 7.5 credits</td>
</tr>
<tr>
<td>Host department:</td>
<td>Department of Cancer Research and Molecular Medicine</td>
</tr>
<tr>
<td>Course coordinator:</td>
<td>Professor Finn Drabløs</td>
</tr>
</tbody>
</table>

**Learning outcome**

After completing the course MOL3022 the student is able to:

- explain the main principles of important algorithms and methods used in bioinformatics tools, including dynamic programming, hidden Markov models and neural networks;
- explain function and usage of important bioinformatics tools, in particular for analyzes at the sequence level (genome, gene, RNA and protein);
- describe important formats for storage and exchange of biological information;
- describe content and use of important bioinformatics databases and web portals;
- use bioinformatics tools and databases to analyze relevant data from molecular biology.

**Academic content**

The course gives a practical and theoretical introduction to important methods in bioinformatics, including sequence library searches, pairwise and multiple alignment, phylogenetic analysis, gene prediction, ontology-based analyses, motif discovery and structure prediction. The students will be able to test the methods on realistic problems through computer-based exercises. The students will also do a method-oriented project highlighting the theory behind some important methods. An individual report
from the implementation of a chosen method will be the basis for grading. The course is in particular targeted towards students with a background from computer science who wants an introduction to important methods that are used to analyse data from molecular biology.

<table>
<thead>
<tr>
<th>MOL3023</th>
<th>Molecular Medical Microbiology with Essay</th>
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<tbody>
<tr>
<td>Credits:</td>
<td>7.5</td>
</tr>
<tr>
<td>Period:</td>
<td>Spring</td>
</tr>
<tr>
<td>Teaching methods:</td>
<td>Lectures and a compulsory written assignment which counts 30 % of the final grade. The language of teaching and examination is English. Alternative examination may be held in case of few students. Timetables for courses at the Faculty of Medicine are available at <a href="https://timeplan.medisin.ntnu.no/timetable_show.php">https://timeplan.medisin.ntnu.no/timetable_show.php</a></td>
</tr>
<tr>
<td>Recommended previous knowledge:</td>
<td>Knowledge in microbiology, cell biology, biochemistry, and/or molecular biology</td>
</tr>
<tr>
<td>Mode of assessment:</td>
<td>4-hour written examination – 70 % of the final grade Assignment – 30 % of the final grade Letter grades (A-F)</td>
</tr>
<tr>
<td>Credit reduction due to overlapping course:</td>
<td>MOL3003: 7.5 credits MOL8011: 7.5 credits</td>
</tr>
<tr>
<td>Host department:</td>
<td>Department of Laboratory Medicine, Children’s and Women’s Health</td>
</tr>
<tr>
<td>Course coordinator:</td>
<td>Professor Jan Egil Afset</td>
</tr>
</tbody>
</table>

**Learning outcome**

After completing the course MOL3023 the student is able to:

- demonstrate a strong general knowledge in molecular medical microbiology;
- have good knowledge of the PCR method including design, optimization and validation for detection, identification and quantification of microorganisms, and be able to interpret PCR results and discuss limitations of the method;
- discuss interpretation of PCR results and limitations of the method;
- demonstrate knowledge of other molecular methods for identification and quantification of microorganisms;
- demonstrate good knowledge of molecular methods for genotyping of microbial agents, and discuss the use of molecular epidemiological methods in the investigation of infectious diseases.

**Academic content**

Molecular methods are important for the diagnosis of infectious diseases and for characterization of the microorganisms causing such infections. In this course the student will be introduced to the use of molecular genetic methods in the diagnosis of human pathogenic microorganisms, including methods for extraction of nucleic acids, qualitative and quantitative PCR and DNA sequencing. Principles for primer and probe design will be explained, as well as use of bioinformatics in the design of PCR assays. Methods for genotyping of microorganisms are important for the study of transmission and dissemination of infectious diseases in hospitals and in the community. A range of genotyping methods developed for this purpose will be presented and discussed.
Learning outcome
After completing the course MOL3100 the student is able to:

- describe the organization of the human genome and its functional regulation (i.e. replication, gene expression, genome maintenance, and signal transduction principles);
- describe the impact of genes, inheritance and environment on disease;
- understand how normal cellular processes change, fail or are destroyed by disease development, in particular for common diseases such as cancer, diabetes, and heart disease;
- collect relevant background information about topics within molecular medicine, use EndNote, and to write a scientific review article of a given topic in English;
- read and present scientific papers.

Academic content
The lectures will cover the organization and major features of the regulation of the function of the human genome, such as gene expression, replication and genome maintenance. Consequences of mutations and polymorphisms, and impacts of genes and environment on major common diseases, such as cancer, diabetes, vascular and coronary disease, will be covered. Basic principles of extracellular and intracellular signalling systems will also be included. Methods of DNA analysis, gene technology and applied bioinformatics will be discussed.

MOL3901 Thesis in Molecular Medicine
Credits: 60
Teaching method: Individual supervision

Admission requirements: The student must be admitted to the Master of Science in Molecular Medicine. In order to be eligible to defend his/her master's thesis the student must have passed all exams, i.e. compulsory and elective courses worth 60 credits in total.

Compulsory activity: Individual supervision
Form of examination: Thesis and oral presentation / examination. The grade given on the thesis may be adjusted after the oral examination.

Host department: Department of Laboratory Medicine, Children’s and Women’s Health
Course coordinator: Associate Professor Wenche Sjursen
Learning outcomes
After successful defense of the thesis the student is able to:

- formulate a precise research problem;
- scientifically test and answer a research problem;
- prepare and analyze data from a study/experiment - master methods and techniques relevant for the research problem;
- present a research problem and discuss the results critically by use of relevant scientific literature;
- describe a scientific work in a clearly written report (master’s thesis);
- present the results, both in writing and orally, with sound language and precise statements.

Academic content
The master's thesis in molecular medicine could have a basal or a medical direction. The thesis should have a scientific composition and be founded on applicable theory and literature within the specific subject.

Current topics could be molecular mechanisms and epidemiological causes, diagnostic problems and therapeutic measures. It is a great advantage that the thesis is connected to the existing research activities at the Faculty of Medicine. The principal supervisor is chosen among the scientific staff with permanent positions.

The thesis should be a monograph or a scientific paper with concluding remarks. In both cases, the student should document a theoretical comprehension and a broad understanding of the methods that have been used. Students enrolled in the master's programme in 2007 or earlier can write the thesis in Norwegian or English. Students enrolled in the master's programme in 2008 or later must write the thesis in English.

Details regulating the work with and assessment of the thesis are given in Guidelines for the Master's Thesis in Molecular Medicine. More information is available at www.ntnu.edu/dmf/studies/master.
Laboratory Animal Science for Researchers

**Credits:** 7.5

**Period:** Autumn

**Teaching methods:** Lectures, study groups and individual assignments. The course consists of 35 hours of lectures (given in week 47), 24 hours of self-stuution (group work and individual home assignments) and 21 hours of practical training. The home assignment must be submitted and passed before written exam can take place. Students who have followed the theoretical lectures, passed the home assignment and passed the written exam will receive a FELASA diploma. This diploma is valid together with a practical training document. The requirements are set by the Department of Agriculture (http://oslovet.veths.no/Oppl/nye.html#KatC). You must arrange the 3 days of practical training yourself. This is usually done in your own research group. The training must be supervised by a person with FELASA C or B competence. Then the practical training documentation must be signed and approved by the local competent person at your laboratory animal facility.

**Recommended previous knowledge:** Biomedical education, courses in statistics, knowledge of literature search on the internet and in the library.

**Required previous knowledge:** A 3-year education on university or college level is a prerequisite in order for the participant to use the title "FELASA category C, Researcher" when the compulsory activities (see the below) have been carried out. Enrolment in a PhD programme, master programme or at "forskerlinjen" in medicine at NTNU. PhD and "forskerlinje" students at the medical faculties at the universities in Bergen, Oslo and Tromsø are given access according to a mutual agreement between these institutions. Others are referred to the course MDV6003.

**Compulsory activities:** Lectures (five days)

**Mode of assessment:**
- 4-hour written examination – 70 % of the final grade
- Report – 30 % of the final grade
- Passed/not passed

**Host department:** Department of Cancer Research and Molecular Medicine

**Course coordinator:** Clinic Veterinarian Siv Eggen

**Learning outcome**

After completing the course NEVR8014 the student:

- shall know the principles behind modern theory on animal experiments and welfare
- knows the legislature regulating the use of lab animals in Norway
- knows the potential health hazards related to animal experiments, and how to minimize these hazards
- understands the significance of the internal and external factors influencing a lab animal and which thereby may influence the outcome of the experiment
- knows roughly how to monitor the health of lab animals
- understands the most important principles for choosing methods for handling and treating lab animals
- understand the principles behind anaesthesia, analgesia and humane killing of lab animals
- understands the general principles for planning animal experiments, including quality control and know of the potential alternatives and supplements to animal experiments which exist
- is able to evaluate a published article on animal experiments with emphasis on how the animals are described and used and know of and be able to use guidelines for good reporting of animal experiments
- has insight into the most important factors which decide the running of a research department using lab animals and be able to do a simple evaluation of a department
has an attitude towards the lab animals which reflect "the three R's" with focus on animal protection and animal welfare (Replace, Reduce, Refine).

**Academic content**

FELASA = Federation of Laboratory Animal Science Associations. We follow their minimum recommendations for education and training for researchers (FELASA C researcher). This means that you most likely can travel with your Diploma (FELASA C) to other European countries and work with Laboratory animals. But other countries might ask for additional training and have more stringent rules. The course will focus on the following topics: Legislation, Ethics and views in society; the course of events in animal experiments; biology of lab animals; the choice of species; genetical and environmental factors influencing animal experiments; health hazards; principles concerning the handling of animals, anesthesia, analgesia and humane killing of lab animals; evaluation and quality control of animal experiments; reporting; alternatives to animals experiments; literature search. The course is divided into two sections; a general section (3 days) and a selectable section (2 days) where the students can choose between traditional laboratory animals and fish/aquatic organisms. Course participants should select their specialization on the basis of the animals they will work with after the course. Traditional animals specialization: laboratory animals biology, health monitoring, anesthesia and euthanasia, ethology, genetics, transgenic animal models, handling techniques. Fish specialization: Legislation concerning fish, experimental conditions, stress, biorythms and acclimatization, pain and suffering, anesthesia, handling, surgical procedures and euthanasia, aggression and hierarchy formation, health monitoring and microbiological qualities, transgene fish.